# The role of nonsteroidal antiinflammatory drugs in cancer chemoprevention

# Charles Y.F. Young

Departments of Urology, and Biochemistry and Molecular Biology, Mayo Clinic College of Medicine, Mayo Clinic/ Foundation, Rochester, Minnesota, 55905, USA; e-mail: young.charles@mayo.edu

### CONTENTS

Abstract	467
Introduction	467
Potential mechanisms for NSAID-mediated cancer	
chemoprevention	468
Cyclooxygenases and carcinogenesis	468
Specific COX-2 inhibitors vs. nonselective	
cyclooxygenase inhibitors	469
NSAIDs and cyclooxygenase in cancer prevention	469
Cyclooxygenase dependence	469
Cyclooxygenase independence	470
Bioactivity and bioavailability of NSAIDs	472
References	473

### **Abstract**

Epidemiological and laboratory studies, as well as human clinical intervention trials, strongly support a protective role for nonsteroidal antiinflammatory drugs (NSAIDs) in preventing the development of cancer. For example, NSAIDs are effective for reducing existing polyps in patients with familial adenomatous polyposis and reduce the tumor burden in several animal models of colorectal cancer. Many studies suggest that cyclooxygenase, especially COX-2, can be causally connected to carcinogenesis via modulation of the synthesis of prostaglandins and other eicosanoids, and subsequent effects on cell proliferation, angiogenesis, tumor growth, immune responsiveness and inflammatory reactions. It is therefore reasonable to assume that NSAIDs as inhibitors of COX-2 would be good candidates for cancer prevention, or even treatment. Animal studies and in vitro cell culture bioassays further demonstrated that cyclooxygenaseindependent actions of NSAIDs could also contribute to their anticancer activities. These studies showed that NSAIDs can affect the expression and functions of many transcription factors related to oncogenesis, including hypoxia-inducible factor-1 (HIF-1), GATA-6, NF-κB, β-catenin/TCF-4, early growth response-1 (Egr-1) and the androgen receptor.

### Introduction

Nonsteroidal antiinflammatory drugs (NSAIDs) have been widely used as antiinflammatory, antipyretic and pain-relieving (or analgesic) medicines, and some can even be purchased for regular use without a prescription. Evidence accumulating over the past three decades strongly suggests that NSAIDs may be useful for protecting from the development and/or progression of several types of cancer, including breast, lung, colorectal, esophageal and prostate cancer. Since cyclooxygenase type 2 (COX-2) has been causally connected to carcinogenesis, it is reasonable to propose that NSAIDs as inhibitors of COX-2 would be good candidates for use in cancer prevention, or even treatment (1-10).

Results from epidemiological studies, laboratory tests and clinical trials strongly support a protective role for NSAIDs against cancer development (11-18). Several epidemiological studies have shown a significant inverse association between the intake of aspirin and other NSAIDs and the risk of colorectal cancer in the general population (3, 11-15). Furthermore, NSAIDs are effective for reducing existing polyps in familial adenomatous polyposis and reduce the tumor burden in several animal models of colorectal cancer.

A study based on an analysis of 1,392 confirmed cases of breast cancer in a population of 80,741 post-menopausal women between the ages of 50 and 79 years, with no history of breast cancer or other cancers, was performed to evaluate breast cancer risk factors and NSAID use (10). The results showed that regular NSAID use for 5-9 years or for 10 years or longer was associated with a reduction of 21% and 28%, respectively, in the incidence of breast cancer. Interestingly, the study showed that the long-term use of ibuprofen provided a greater risk reduction than aspirin, whereas acetaminophen or low-dose aspirin (< 100 mg) was not effective in reducing the incidence of breast cancer.

Esophageal carcinomas are associated with a high mortality rate, making chemopreventive agents desirable. A meta-analysis of 9 studies (2 cohort, 7 case-control) including 1,813 cases assessed the relationship between aspirin/NSAID use and esophageal cancer (9). The

findings showed that both aspirin and nonaspirin NSAIDs had protective effects against esophageal adenocarcinoma and squamous cell carcinoma, more frequent use producing greater protection.

A similar inverse association has been described for other cancers, including ovarian and lung cancers. For prostate cancer, there are a few studies showing a weak, inverse association (not statistically significant) between NSAID use and cancer (19, 20). However, a recent report by Nelson and Harris (21) indicated a 70% reduction in the risk of prostate cancer among NSAID users. A more recent, community population-based study (18) at the Mayo Clinic showed a significant reduction in prostate cancer risk among men aged 60 or older who were daily users of NSAIDs. Interestingly, the study found the strongest inverse association among men aged 70 or older, with an 83% reduction in risk. The authors attributed this to the greater use of NSAIDs in this group of men.

# Potential mechanisms for NSAID-mediated cancer chemoprevention

Cyclooxygenases and carcinogenesis

Although the mechanism(s) by which NSAIDs exert anticancer activity are far from completely understood, increasing evidence indicates that inhibition of arachidonic acid (AA) metabolic pathways via cyclooxygenase may contribute to the anticancer activity of NSAIDs (1-10). Cyclooxygenases can modulate the synthesis of prostaglandins and other eicosanoids that affect cell proliferation, angiogenesis, tumor growth, immune responsiveness and inflammatory reactions (11-12, 14, 15). Cyclooxygenase enzymes may be involved in activation of carcinogens and the formation of reactive oxygen species (11). Thus, NSAIDs that inhibit cyclooxygenase enzyme activities may be potential chemopreventive agents.

There are two cyclooxygenase genes, COX-1 and COX-2, located on chromosomes 9 and 1, respectively; COX-1 is constitutively expressed in many tissues for normal physiological functions and COX-2 is inducible by cytokines in mediating inflammation (3, 11). Both COX-1 and COX-2 share about 60% homology in amino acid sequence. The active site of these isozymes only differs at two positions. In fact, both isozymes have almost identical catalytic kinetics, causing oxygenation of arachidonic acid to produce PGG, and PGH, and subsequent prostaglandins and thromboxane by separate enzymes depending on cell or tissue type. However, COX-2 may be able to use more alternative substrates than COX-1, e.g., COX-2 can use docosahexaenoic acid and endocannabinoids as substrates. As mentioned above, both COX-1 and COX-2 have the same substrate specificity, and specific inhibitors with clinically demonstrable selectivity for the isozymes have been developed and marketed.

Interestingly, the finding that the potent antipyretic and analgesic effects of acetaminophen could not be explained by either COX-1 or COX-2 blockade eventually led to the discovery of COX-3, a splice variant of COX-1, in canine brain (22-24). The COX-3 variant is sensitive to acetaminophen and considered to play a critical role in the biosynthesis of prostanoids known to be important mediators in pain and fever. Drugs that preferentially block COX-1 also appear to act at COX-3. In addition, other splice variants of COX-1 have been described and may be detectable in human tissues, although their significance requires further study. Moreover, the existence of COX-3 at the nucleotide sequence level in humans has not been firmly proven.

Several lines of evidence have been presented that link cyclooxygenase and its prostaglandin products to cancer development, and perhaps progression. The first clue was that cancer tissue prostaglandin levels are often correlated with poor postoperative survival or the development of metastasis (25, 26). Increased levels of prostaglandins, especially PGE2, were found in many cancer tissues compared to their normal counterparts (27-31). Similar results were also found in cancer tissues from experimental animal models of carcinogenesis. It has been shown that PGE2 can be synthesized in cancer cell lines. In cell culture systems, PGE2 may enhance tumor cell proliferation.

In the early 1990s, COX-2 was shown to be one of the proteins induced by the viral oncogene c-src in cell transformation or by growth factors/receptors, including the EGF (epidermal growth factor) receptor, the IGF-1 (insulin-like growth factor) receptor, the heregulin/HER-2 receptor or mutations in APC and ras (6, 8, 32, 33). The COX-2 isozyme has been shown to be overexpressed in many tumor tissues and cancer cell lines (27-31). Overexpression of COX-2 via transfection was associated with increases in PGE, and Bcl-2, and a subsequent decrease in apoptosis (8). Recently, a transgenic model demonstrated that overexpression of COX-2 alone is enough to cause mammary tumorigenesis. Furthermore, chemically induced mammary tumorigenesis in rats can be prevented by the selective COX-2 inhibitor celecoxib. It thus appears that COX-2 may be more relevant than COX-1 to tumor development.

However, some studies indicated that COX-1 could also have a role in carcinogenesis. COX-1 knockout Min mice showed an 80% reduction in the development of polyposis (34). A more recent study (35) seemed to suggest that COX-1 constitutively expressed in stromal fibroblasts and providing basal levels of PGE<sub>2</sub> could be critical for the initial establishment of polyps. In both animal and human tissues, COX-1 and COX-2 were found to be co-expressed in fibroblast cells in polyps, although the expression of COX-2 is induced when polyps grow to over 1 mm in size. COX-2 may produce more PGE<sub>2</sub> for boosting tumor growth. Furthermore, as suggested in some studies, one intriguing aspect is that increasing COX-2 activities with increased PGE<sub>2</sub> may be related to immunosuppressed states of cancer, resulting from

increasing IL-10 and decreasing IL-1, IL-12 and tumor necrosis factor (TNF) levels (8). Of course, the inhibition of cyclooxygenase by NSAIDs resulting in reduction in cancer risk would prove the action of cyclooxygenase in carcinogenesis.

Furthermore, COX-2 can play an important role in tumor angiogenesis and invasion by modulating the expression and function of vascular endothelial growth factor (VEGF) and integrin-mediated signaling via COX-2 enzymatic products (1, 6, 8). Angiogenesis is a hallmark of tumor expansion and metastasis. In the early 1970s, it was suggested that the new capillary blood vessels (or microvessels) are necessary for cancer cells to grow if tumor mass reaches 1-2 mm in diameter, because at that point the supply of the nutrients, including oxygen, can not reach those tumor cells distant from the nearest vessels. Only those tumors capable of stimulating and developing new microvessels may have the potential to become metastatic.

There are two types of cells, *i.e.*, endothelial cells and pericytes, in vessels that can respond to proper stimuli to form new vessels. Normally these cells are quiescent, with a life span of several years. However, many angiogenic or proangiogenic agents can stimulate these cells, including those from tumor cells. Once the proangiogenic growth factors diffuse from tumor cells to preexisting blood vessels, quiescent enthothelial cells will be activated and induce a cascade of events, including proteolytic breakdown of the basement membrane, migration of the endothelial cells towards the angiogenic sources and proliferation of the cells, lumen formation, pericyte capping, and finally, formation of a new basement membrane for the new microvessels.

Cyclooxygenases, especially COX-2, have been suggested to be directly related to the angiogenesis of tumors. As mentioned above, catalytic activities of increased COX-2 will enhance the production of eicosanoids, including thromboxane A2 (TxA2), PGE2 and PGI<sub>2</sub>, which are essential for neovascular formation, providing sufficient nutrients for subsequent tumor growth. Thromboxane A2 also plays a critical role in COX-2dependent endothelial migration and neovascularization of tumors (1, 6, 8). It has been shown that migration of the endothelial cells can be inhibited by a COX-2-selective inhibitor via a reduction in TxA2 levels. Adding exogenous TxA2 to the cells can reverse the effects of the COX-2selective inhibitor. Overexpression of COX-2 appears to be responsible for the production of many proangiogenic molecules, such as VEGF, basic fibroblast growth factor (bFGF), transforming growth factor- $\beta$  (TGF- $\beta$ ) and platelet-derived growth factor (PDGF), as well as integrinmediated signaling (1, 6, 8). Metalloproteinases may be upregulated by COX-2 via PGE, in prostate and colon cancer cell lines.

Prostaglandins as cyclooxygenase metabolic products exert their biological actions through binding to their plasma membrane receptors (prostanoid receptors) with seven transmembrane domains (36). Prostaglandin-activated receptors appear to act via G-protein to cAMP and protein kinase A to increase VEGF. Studies using prostanoid receptor knockout mice strongly suggested that the prostanoid receptor signal is important for tumorassociated angiogenesis and subsequent tumor growth. Experiments in prostanoid receptor knockout mice bearing tumor xenografts showed that receptors in the host stroma could play a critical role in tumor-associated angiogenesis. The studies indicated that antagonizing prostanoid receptors on host stromal cells is more effective than inhibiting the receptors in tumors for preventing tumor growth. Developing antagonists for prostanoid receptors may therefore represent a new approach to cancer prevention.

Specific COX-2 inhibitors vs. nonselective cyclooxygenase inhibitors

Chronic use of traditional NSAIDs which block both cyclooxygenase enzymes may cause serious side effects such as gastrointestinal bleeding, ulceration and renal toxicity (3, 11, 14, 15). The side effects of traditional NSAIDs are due mainly to inhibition of COX-1. As mentioned above, COX-1 is generally constitutively expressed in many tissues, whereas COX-2 is inducible by proinflammatory cytokines (3, 11). Importantly, COX-2 has been found to be overexpressed in many tumor tissues (3, 15). Therefore, improved, new-generation NSAIDs are designed to target COX-2 and thereby reduce the above side effects (3, 14, 15).

Large randomized trials have recently been performed evaluating long-term use of NSAIDs, including the Vioxx Gastrointestinal Outcome Research and the Celecoxib Long-Term Arthritis Safety Study (CLASS) (37, 38). These studies appeared to show that the COX-2selective NSAIDs are associated with an improved gastrointestinal safety profile compared to the classical NSAIDs. However, a cautionary note has been raised in a study regarding the potential cardiovascular risk associated with the use of the NSAIDs (39). Another unexpected result from more recent studies suggested that aspirin might increase the risk of human pancreatic cancer (40, 41), whereas previous studies demonstrated that NSAIDs can decrease experimental pancreatic carcinogenesis in hamsters (42). A previous human survey also showed that frequent use of aspirin may reduce the risk of pancreatic cancer (43). In vitro data indicated antineoplastic effects for NSAIDs on pancreatic cells (44). However, the latest study (40) suggested that animal models and in vitro studies may not be very relevant to humans. Clearly, more studies should be performed to clarify the issue.

# NSAIDs and cyclooxygenase in cancer prevention

Cyclooxygenase dependence

Recently, it has been proposed that activation of inducible nitric oxide synthase (iNOS) and consequent

generation of NO may be associated with the development of cancer in a variety of gastrointestinal diseases. Furthermore, both COX-2 and iNOS have been reported to be implicated in breast and colon cancer progression (45, 46). Interestingly, another study showed that PGE<sub>2</sub> generated from COX-2 expression in the highly metastatic murine breast cancer cell line C3L5 can upregulate interferon gamma + lipopolysaccharide (LPS)-induced iNOS expression and NO production (47). It was further demonstrated that upregulation of iNOS by PGE<sub>2</sub> is mediated by the PGE<sub>2</sub> EP<sub>4</sub> receptor in a cAMP-dependent manner. The increasing levels of NO can be suppressed by NSAIDs, which is reversible by exogenous PGE<sub>2</sub> or the EP<sub>4</sub> receptor agonist PGE<sub>4</sub> alcohol.

As discussed above, it is generally agreed that the chemopreventive function of NSAIDs may be due to their cyclooxygenase-inhibitory effect on prostaglandin production. Furthermore, it was reported that NSAIDs such as sulindac and indomethacin could induce apoptosis in colon cancer cells by accumulating a relative high concentration of arachidonic acid, which in turn stimulated the conversion of sphingomyelin to ceramide, a known mediator of apoptosis (48). Accumulation of arachidonic acid by inhibiting its conversion to PGE, can therefore activate sphingomyelinase, leading to an increase in ceramide. This effect may not be restricted to colon cancer cells. It could also be observed in primary fibroblasts and immortalized keratinocytes. Thus, this effect might have general application to NSAID-mediated cancer prevention.

It has been known for many years that high serum estrogen levels may increase the risk of breast cancer (2, 49-52). Recently it was shown that aromatase (CYP19) can be detected in breast tumor tissues and may catalyze the production of estrogens in the tumor tissues (53-55). Intratumor aromatase and estrogen production may have a role in breast cancer development and progression. The expression of aromatase in normal and cancerous breast tissues may be differentially regulated. A so-called promoter switch theory has been proposed in which cAMP may be a critical factor causing alternative promoter activation in the aromatase gene. PGE<sub>2</sub> produced by COX-2 and certain cytokines can upregulate the expression of aromatase (56, 57). Interestingly, it was demonstrated using a semiguantitative RT-PCR assay (58) that COX-2 and aromatase were both overexpressed in 23 human breast cancer tissues. This finding seems to indicate that selective NSAIDs targeting COX-2 to reduce PGE, and, subsequently, aromatase activity would be an appealing strategy for reducing breast cancer risk, and potentially other hormonerelated cancers.

Several transcription factors can be regulated by NSAIDs. These factors may be related to cell proliferation and angiogenesis. In ischemic conditions, the HIF complex is activated and binds a hypoxia response element in the promoters of the VEGF gene and the VEGF receptor Flt-1 (59-66). The HIF complex consists of a heterodimer of HIF-1 $\alpha$  and an aryl hydrocarbon receptor nuclear

translocator (HIF-1β). Under normoxic conditions, HIF-1β is relatively stable, but HIF-1 $\alpha$  binds to the von Hippel-Lindau tumor suppressor protein (VHL) and becomes susceptible to rapid degradation via ubiquitination. On the other hand, under hypoxia, HIF-1 $\alpha$  and HIF-1 $\beta$  form a stable complex and bind a hypoxia response element in the promoters of the VEGF and Flt-1 genes onto which AP-1 proteins are recruited, followed by the initiation of transcription. It is known that hypoxia by over- and outgrown tumor tissue mass may trigger neovascularization in the tumor mass and help tumor cells grow and spread. NSAIDs can inhibit hypoxia-induced angiogenesis. Although it has been shown that NSAIDs decrease VEGF and Flt-1 by reducing HIF-1 expression via an increase in VHL levels and an increase in ubiquitination of HIF-1 $\alpha$ , exactly how NSAIDs induce downregulation of HIF-1 is not clear. Since  $PGE_2$  was shown to induce  $HIF-1\alpha$  stability and nuclear localization, it is reasonable to assume that inhibition of PGE, production by NSAIDs may contribute to the downregulation of HIF-1α and its downstream products VEGF and Flt-1 (59-66).

## Cyclooxygenase independence

The early growth response factor Egr-1 is another transcription factor that is important in angiogenesis induced by hypoxia (67-69). Egr-1 can regulate a number of genes, including Flt-1 and epidermal growth factor (EGF) receptor. Vascular endothelial growth factor can induce Egr-1 expression, which in turn activates the expression of Flt-1 (68-72). Therefore, as mentioned above, VEGF expression could be affected by NSAIDs via inhibition of cyclooxygenase activity. In one study (73), NSAIDs such as indomethacin and the COX-2 inhibitor NS-398 were shown to be able to inhibit Egr-1 expression stimulated by exogenous VEGF in cultured human microvascular endothelial cells (HMVEC). Since the effective concentrations of the NSAIDs used to inhibit the expression of Egr-1 were much higher than those for repressing cyclooxygenase activity, the effect of NSAIDs in this case might be viewed as a cyclooxygenase-independent mechanism.

The APC/ $\beta$ -catenin/TCF pathway (Wnt signaling pathway) has been suggested to be involved in the initial stages of the adenoma-carcinoma sequence. Treatment with sulindac of 5 patients with familial adenomatous polyposis (FAP) for up to 6 months was found to reduce adenoma nuclear  $\beta$ -catenin expression when compared to pretreatment adenomas from the same patients (74). In vitro studies (75, 76) with sulindac sulfide, a sulindac metabolite lacking cyclooxygenase-inhibitory activity, in the colorectal cancer cell lines DLD-1 and SW480 showed decreased levels of nonphosphorylated  $\beta$ -catenin and a subsequent reduction in  $\beta$ -catenin-mediated transcription of the target genes Met and cyclin D1. This action of sulindac sulfide suggests a COX-2-independent mechanism in the chemopreventive effect of NSAIDs.

Moreover, several mechanisms have been proposed regarding how NSAIDs affect the transcriptional

function of  $\beta$ -catenin/TCF (76). For example, the downregulation of  $\beta$ -catenin may be caused by caspase-and proteasome-dependent degradation pathways, as a result of its enhanced phosphorylation (77) by glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ) in SW948 and SW480 colorectal cancer cells (78, 79). On the other hand, it was shown that both aspirin and indomethacin can inhibit  $\beta$ -catenin/TCF-mediated transcription without affecting its levels (78, 79). In an animal study, sulindac, indomethacin, meloxicam and sulindac sulfone all effectively suppressed chemically induced rodent colorectal tumor formation and inhibited  $\beta$ -catenin nuclear translocation as assayed by immunohistochemical staining (75).

Interestingly, a new class of NSAID, *i.e.*, NO-donating aspirin, has been developed recently with much higher potency than the parent aspirin (80-82). This new chemical can inhibit NOS2 expression/NO formation,  $\beta$ -catenin/TCF-mediated transcriptional activity and NF- $\kappa$ B DNA binding ability at concentrations lower than or near those required for inhibiting cell growth or killing cells. Thus, NO-aspirin may act at critical early events in tumor formation as a chemopreventive agent in colon cancer.

It has been shown that NSAIDs can suppress cancer cell proliferation, soft agar colony formation and the induction of apoptosis regardless of the cyclooxygenase status of the cells (3, 83-85). Sulindac sulfone, a metabolite of the NSAID sulindac that lacks cyclooxygenaseinhibitory activity, blocked tumor growth in mice (16, 86). Moreover, sulindac, an inhibitor of COX-1 and COX-2, induced apoptosis by blocking the antiapoptotic protein Bcl-X and inactivating peroxisome proliferator-activated receptor PPARδ. Also, celecoxib was reported to downregulate the antiapoptotic protein Bcl-2 (85). However, another study (87) reported that the apoptosis induced was due to blockade of Akt activation independent of Bcl-2. Furthermore, aspirin, sulindac and sulindac sulfone all inhibited tumor cell proliferation by blocking activation of I-κB kinase- (IKK-β), which is required for activation of NF-κB (88). Active NF-κB enhances cell survival.

Moreover, NSAIDs can inhibit cyclooxygenase-independent angiogenesis (92). Recently, it has been shown (90) that sulindac downregulates the transcription repressor GATA-6 for the 15-lipoxygenase-1 (15-LOX-1) promoter and enhances the expression of 15-LOX-1, as well as the product of 15-LOX-1, 13-(S)-hydroxyoctadecadienoic acid (13-[S]-HODE), which in turn enhances the apoptosis of colorectal cancer cells. It has been shown that linoleic and arachidonic acid metabolites can be peroxisome proliferator-activated receptor (PPAR) ligands and the PPARS can enhance cell survival and proliferation, and promote colonic tumorigenesis (91-96). Moreover, activated PPARδ can upregulate the expression of COX-2 (97-99). It was shown that 13-(S)-HODE can act as a PPARS antagonist and decrease PPARS activation, subsequently downregulating PPARδ expression and resulting in apoptosis in colorectal cancer cells. These studies seem to provide a mechanistic link through NSAID-induced 15-LOX-1 and its product 13-(S)-HODE to induction of apoptosis by downregulation of PPARδ. It is clear that NSAIDs possess cyclooxygenase-independent anticancer effects. However, *in vitro* experimental results should be interpreted cautiously, because many studies did not clearly define or can not judge whether the actions of the NSAIDs used are strictly cyclooxygenase-independent.

Recently, we found that NSAIDs may exhibit a new biological activity: suppressing the androgen receptor in prostate cancer cells (100, 101). Prostate cancer exceeds lung cancer as the most commonly diagnosed cancer in males and is the second leading cause of male cancer death in many western countries. The androgen receptor, a ligand-dependent transcription factor, plays a central role in androgen actions in the prostate, as well as being a risk factor for the development and progression of prostate cancer (102-109). A prospective cohort study demonstrated a strong positive association between plasma testosterone levels and prostate cancer risk (107). Removal or blockade of the synthesis of androgens, thereby inactivating androgen receptor activity, can result in the induction of apoptosis in both normal and cancerous prostatic epithelia in vivo (102, 110, 111). Males with 5α-reductase deficiency or castration at an early age do not develop benign or malignant prostate tumors (102, 103, 111). The above evidence provides strong support for androgen deprivation-based chemoprevention and chemotherapy for prostate neoplasia.

The androgen receptor is considered to be the central factor in the development and progression of prostate cancer. However, it has not clearly been explained how prostate cells are transformed and become malignant, although it is thought that growth stimulation (proliferation) can increase the number of mutated cells, which are further stabilized (survival) by androgens and their receptor. The transformation to malignancy is mainly induced by genetic changes in normal cells via environmental factors (mutagenic factors: chemical and physical) and endogenous factors (hormones, growth factors, etc.) (112-114). Recently, it was postulated that androgens and the androgen receptor could be potentially promutagenic. Overactivation of the androgen receptor due to high levels of androgens might produce excessive reactive oxygen species (ROS), which can be mutagenic and harmful (115, 116). The interaction with other environmental factors (dietary factors such as animal fat) can further enhance the transformation rates of prostate cells in vivo. Recently, overexpression of many cofactors (e.g., caveolin-1, STAT3, β-catenin) or coactivators (e.g., TIF2 and SRC-1) for the androgen receptor was suggested to enhance the function of the receptor at low levels of androgens or in the presence of other nonandrogen ligands (117-129). Of course, mutation and amplification of the androgen receptor can also enhance the function of the receptor in a similar way. Additionally, polymorphism in a trinucleotide repeat (CAG) encoding a polyglutamine stretch in the N-terminal domain of the androgen receptor has been associated with an increasing risk of prostate cancer (if the CAG tract is shortened) (130-132).

Although epidemiological studies suggest a negative association between consumption of NSAIDs and the risk

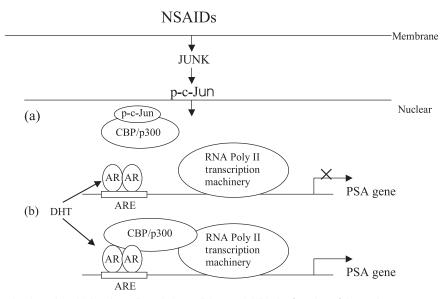


Fig. 1. Proposed mechanisms by which celecoxib and nimesulide may inhibit the function of the androgen receptor (AR) in prostate cells. (a) After treatment with the NSAIDs, JUNK is activated, which may cause phosphorylation of c-Jun (p-c-Jun) and consequently prolong overexpression of c-Jun protein. Prolonged overexpression of c-Jun could in turn sequester the coactivator CBP/p300 from the AR, thereby inhibiting its function even in the presence of androgens such as dihydrotestosterone (DHT). (b) Upon binding and activation by androgens such as DHT, the AR in the nucleus recruits CBP/p300 while binding to the androgen response element (ARE) of the promoter of androgen-regulated genes, including the prostate-specific antigen (PSA) gene. CBP/p300 can acetylate the AR and enhance its activity, and also acts as an integrator between the AR and basic transcription machinery for activation of the expression of androgen-regulated genes.

of prostate cancer, the COX-2 protein or its enzymatic activity is not frequently overexpressed in prostate cancer tissues (91, 133). This suggests that cyclooxygenase inhibition by NSAIDs may not be the sole mechanism for prostate cancer prevention. According to our recent studies, the COX-2 inhibitors celecoxib and nimesulide appear to have much higher potency than several other NSAIDs tested in inhibiting the expression and/or function of the androgen receptor. Thus, their *in vivo* effective concentrations may be achievable, in contrast to other NSAIDs. Modulating the androgen receptor may be one of the mechanisms by which these NSAIDs can inhibit the development of prostate cancer. This indicates that these two NSAIDs, and perhaps also their analogues, may be potential agents for prostate cancer prevention.

Our studies indicated that the NSAIDs could stimulate a prolonged overexpression of c-Jun (101). It was suspected that, in part, phosphorylation of c-Jun could stabilize and therefore prolong the expression of c-Jun. c-Jun is one of the components of activating protein-1 (AP-1) and a member of the basic leucine zipper (bZIP) family of sequence-specific dimeric DNA-binding proteins. It has been shown that Jun can control a highly diverse set of genes, many of which may be related to cellular proliferation processes (134-136). Recently, it was also demonstrated that Jun proteins may be related to cellular apoptosis and growth inhibition. Interestingly, c-Jun can act as a positive transcription factor or a repressor. It can bind CBP/p300, SRC-1 or TIF2/GRIP1 and other coactivators as a positive transcription factor (137-141). On the other

hand, it can bind to steroid nuclear receptors and act as a repressor (137, 138). In different cases, c-Jun can interact with corepressors such as N-CoR, SMRT or TG-interacting factor to repress gene expression (137-141).

It has been shown that members of the AP-1 protein family are upregulated in the castrated prostate (142, 143). Previous results have shown that stimulated overexpression of c-Jun protein can inhibit the function of the androgen receptor (144-149). Several studies have shown that the transactivation functions of the androgen receptor, as well as other steroid receptors, can be affected by c-Jun (150-154). It was further demonstrated that there is a direct protein-protein interaction between the DNA- and ligand-binding domains of the androgen receptor and the leucine zipper region of c-Jun that might affect the function of the DNA-binding domain of the androgen receptor (153). Additional studies indicated that the transcriptional interference between the androgen receptor and AP-1 is partly mediated through competition for intracellular CBP/p300 (152, 154); CBP serves as an integrator/coactivator for androgen receptor transactivation activities. Further studies are required to ascertain whether c-Jun is indeed able to interfere with the interaction of CBP and the androgen receptor by NSAIDs (Fig. 1).

## Bioactivity and bioavailability of NSAIDs

There are sufficient data demonstrating that the potential in vivo antitumor activities of NSAIDs are

separable from cyclooxygenase-inhibitory activity (155-158). However, many in vitro cell culture systems used to show antiproliferative or apoptosis-inducing activity of NSAIDs require higher concentrations than those inhibiting cyclooxygenase activity (158). The differences in the concentrations for these two different activities of the same NSAIDs may reach 100- to 1,000-fold. Usually, low micromolar concentrations of many NSAIDs are associated with inhibition of cyclooxygenase activity, whereas much higher concentrations of the same NSAIDs are required for in vitro cell growth inhibition or the induction of apoptosis. The high concentrations of NSAIDs could be toxic in vivo and may not be reached in the bloodstream, which could be a necessary safeguard. Few NSAIDs may be able to reach their antiproliferative dose in the circulation.

### References

- 1. Tarnawski, A.S., Jones, M.K. *Inhibition of angiogenesis by NSAIDs: Molecular mechanisms and clinical implications.* J Mol Med 2003, 81: 627-36.
- 2. Davies, G.L.S. *Cyclooxygenase-2 and chemoprevention of breast cancer.* J Steroid Biochem Mol Biol 2003, 86: 495-9.
- 3. Bakhle, Y.S. COX-2 and cancer: A new approach to an old problem. Brit J Pharmacol 2001, 134: 1137-50.
- 4. Hawk, E.T., Viner, J.L., Dannenberg, A., DuBois, R.N. *COX-2 in cancer A player that's defining the rules*. J Natl Cancer Inst 2002, 94: 545-6.
- 5. Majima, M., Amano, H., Hayashi, I. *Prostanoid receptor sig-naling relevant to tumor growth and angiogenesis*. Trends Pharmacol Sci 2003, 24: 524-9.
- 6. Rao, M., Yang, W., Seifalian, A.M, Winslet, M.C. *Role of cyclooxygenase-2 in the angiogenesis of colorectal cancer.* Intl J Colorectal Dis 2004, 19: 1-11.
- 7. Jolly, K., Cheng, K.K., Langman, M.J.S. NSAIDs and gastrointestinal cancer prevention. Drugs 2002, 62: 945-56.
- 8. Romano, M., Claria, J. *Cyclooxygenase-2 and 5-lipoxygenase converging functions on cell proliferation and tumor angiogenesis: Implications for cancer therapy.* FASEB J 2003, 17: 1986-95.
- 9. Corley, D.A., Kerlikowske, K., Verma, R., Buffler, P. *Protective association of aspirin/NSAIDs and esophageal cancer: A systematic review and meta-analysis.* Gastroenterology 2003, 124: 47-56.
- 10. Harris, R.E., Chlebowski, R.T., Jackson, R.D., Frid, D.J., Ascenseo, J.L., Anderson, G., Loar, A., Rodabough, R.J., White, E., McTiernan, A. *Breast cancer and nonsteroidal anti-inflammatory drugs: Prospective results from the women's health initiative.* Cancer Res 2003, 63: 6096-101.
- 11. Levy, G.N. *Prostaglandin H synthases, nonsteroidal anti-inflammatory drugs, and colon cancer.* FASEB J 1997, 11: 234-47.
- 12. Vainio, H. *Is COX-2 inhibition a panacea for cancer prevention?* Intl J Cancer 2001, 94: 613-4.

13. Khuder, S.A., Mutgi, A.B. Breast cancer and NSAID use: A meta-analysis. Brit J Cancer 2001, 84: 1188-92.

- 14. Husain, S.S., Szabo, I.L., Tarnawski, A.S. *NSAID inhibition of GI cancer growth: Clinical implications and molecular mechanisms of action*. Am J Gastroenterol 2002, 97: 542-53.
- 15. Bucher, C., Jordan, P., Nickeleit, V., Torhorst, J., Mihatsch, M.J. Relative risk of malignant tumors in analgesic abusers: Effects of long-term intake of aspirin. Clin Nephrol 1999, 51: 67-72.
- 16. Goluboff, E.T., Shabsigh, A., Saidi, J.A., Weinstein, I.B., Mitra, N., Heitjan, D., Piazza, G.A., Pamukcu, R., Buttyan, R., Olsson, C.A. *Exsulind (sulindac sulfone) suppresses growth of human prostate cancer in a nude mouse xenograft model by increasing apoptosis.* Urology 1999, 53: 440-5
- 17. Myers, C., Koki, A., Pamukcu, R., Wechter, W., Padley, R.J. *Proapoptotic anti-inflammatory drugs.* Urology 2001, 57(4a, Suppl. S): 73-6.
- 18. Roberts, R.O., Jacobson, D.J., Girman, C.J., Rhodes, T., Lieber, M.M., Jacobsen, S.J. *A population-based study of daily nonsteroidal anti-inflammatory drug use and prostate cancer.* Mayo Clinic Proc 2002, 77: 219-25.
- 19. Norrish, A.E, Jackson, R.T, McRae, C.U. *Non-steroidal anti-inflammatory drugs and prostate cancer progression*. Intl J Cancer 1998, 77: 511-5.
- 20. Paganini-Hill, A, Chao, A, Ross, R.K, Henderson, B.E. *Aspirin use and chronic diseases: A cohort study of the elderly.* Brit Med J (BMJ) 1989, 299: 1247-50.
- 21. Nelson, J.E., Harris, R.E. *Inverse association of prostate cancer and non-steroidal anti-inflammatory drugs (NSAIDs): Results of a case-control study.* Oncol Rep 2000, 7: 169-70.
- 22. Chandrasekharan, N.V., Dai, H., Roos, K.L.T., Evanson, N.K., Tomsik, J., Elton, T.S., Simmons, D.L. *COX-3, a cyclooxy-genase-1 variant inhibited by acetaminophen and other anal-gesic/antipyretic drugs: Cloning, structure, and expression.* Proc Natl Acad Sci USA 2002, 99: 13926-31.
- 23. Schwab, J.M,. Schluesener, H.J., Meyermann, R., Serhan, C.N. *COX-3 the enzyme and the concept: Steps towards highly specialized pathways and precision therapeutics?* Prostagland Leuk Essent Fatty Acids 2003, 69: 339-43.
- 24. Simmons, D.L. *Variants of cyclooxygenase-1 and their roles in medicine*. Thromb Res 2003, 110: 265-8.
- 25. Rao, C.V., Simi, B., Wynn, T.T., Garr, K., Reddy, B.S. Modulating effect of amount and types of dietary fat on colonic mucosal phospholipase A<sub>2</sub>, phosphatidylinositol-specific phospholipase C activities and cyclooxygenase metabolite formation during different stages of colon tumor promotion in male F34 rats. Cancer Res 1996, 56: 532-7.
- 26. Eberhart, C.E., Coffey, R.J., Radhika, A., Giardello, F.M., Ferrenbach, S., DuBois, R.N. *Up-regulation of cyclooxygenase 2 gene expression in human colorectal adenomas and adenocar-cinomas*. Gastroenterology 1994, 107: 1183-8.
- 27. Kutchera, W., Jones, D., Matsumani, N., Groden, J., McIntyre, T.M., Zimmerman, G.A., White, R.L., Prescott, S.M. *Prostaglandin H synthase 2 is expressed abnormally in human colon cancer: Evidence for a transcriptional effect.* Proc Natl Acad Sci USA 1996, 93: 4816-20.

- 28. Muller-Decker, K., Albert, C., Lukanov, T., Winde, G., Marks, F., Furstenberger, G. *Cellular localization of cyclooxygenase isozymes in Crohn's disease and colorectal cancer.* Intl J Colorectal Dis 1999, 14: 212-8.
- 29. Gustafson-Svard, C., Lilja, I., Hallbook, O., Sjodahl, R. *Cyclooxygenase-1 and cyclooxygenase-2 gene expression in human colorectal adenocarcinomas and in azoxymethane induced colonic tumours in rats.* Gut 1996, 38: 79-84.
- 30. DuBois, R.N., Shao, J., Tsujii, M., Sheng, H., Beauchamp, R.D. *G1 delay in cells overexpressing prostaglandin endoperoxide synthase-2.* Cancer Res 1996, 56: 733-7.
- 31. Sano, H., Kawahito, Y., Wilder, R.L., Hashiramoto, A., Mukai, S., Asai, K., Kimura, S., Kato, H., Kondo, M., Hla, T. *Expression of cyclooxygenase-1 and -2 in human colorectal cancer.* Cancer Res 1995, 55: 3785-9.
- 32. Vadlamudi, R., Mandal, M., Adam, L., Steinbach, G., Mendelsohn, J., Kumar, R. *Regulation of cyclooxygenase-2 pathway by HER-2 receptor.* Oncogene 1999, 18: 305-14.
- 33. Murono, S., Inoue, H., Tanabe, T., Joab, I., Yoshizaki, T., Furukawa, M., Pagano, J.S. *Induction of cyclooxygenase-2 by Epstein-Barr virus latent membrane protein 1 is involved in vascular endothelial growth factor production in nasopharyngeal carcinoma cells.* Proc Natl Acad Sci USA 2001, 98: 6905-10.
- 34. Chulada, P.C., Thompson, M.B., Mahler, J.F., Doyle, C.M., Gaul, B.W., Lee, C., Tiano, H.F., Morham, S.G., Smithies, O., Langenbach, R. *Genetic disruption of Ptgs-1, as well as of Ptgs-2, reduces intestinal tumorigenesis in Min mice.* Cancer Res 2000, 60: 4705-8.
- 35. Takeda, H, Sonoshita, M., Oshima, H., Sugihara, K., Chulada, P.C., Langenbach, R., Oshima, M., Taketo, M.M. Cooperation of cyclooxygenase 1 and cyclooxygenase 2 in intestinal polyposis. Cancer Res 2003, 63: 4872-7.
- 36. Majima, M., Amano, H., Hayashi, I. *Prostanoid receptor signaling relevant to tumor growth and angiogenesis*. Trends Pharmacol Sci 2003, 24: 524-9.
- 37. Oviedo, J.A., Wolfe, M.M. Gastroprotection by coxibs: What do the Celecoxib Long-Term Arthritis Safety Study and the Vioxx Gastrointestinal Outcomes Research trial tell us? Rheum Dis Clin North Am 2003, 29: 769.
- 38. Silverstein, F.E., Faich, G., Goldstein, J.L., Simon, L.S., Pincus, T., Whelton, A., Makuch, R., Eisen, G., Agarwal, N.M., Stenson, W.F., Burr, A.M., Zhao, W.W., Kent, J.D., Lefkowith, J.B., Verburg, K.M., Geis, G.S. Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis The CLASS study: A randomized controlled trial. JAMA J Am Med Assoc 2000, 284: 1247-55.
- 39. Mukherjee, D., Nissen, S.E., Topol, E.J. *Risk of cardiovascular events associated with selective COX-2 inhibitors.* JAMA J Am Med Assoc 2001, 286: 954-9.
- 40. Schernhammer, E.S., Kang, J.H., Chan, A.T., Michaud, D.S., Skinner, H.G., Giovannucci, E., Colditz, G.A., Fuchs, C.S. *A prospective study of aspirin use and the risk of pancreatic cancer in women.* J Natl Cancer Inst 2004, 96: 22-8.
- 41. Baron, J.A. What now for aspirin and cancer prevention? J Natl Cancer Inst 2004, 96: 4-5.
- 42. Anderson, K.E., Johnson, T.W., Lazovich, D., Folsom, A.R. Association between nonsteroidal anti-inflammatory drug use

- and the incidence of pancreatic cancer. J Natl Cancer Inst 2002, 94: 1168-71.
- 43. Kokawa, A., Kondo, H., Gotoda, T., Ono, H., Saito, D., Nakadaira, S., Kosuge, T., Yoshida, S. *Increased expression of cyclooxygenase-2 in human pancreatic neoplasms and potential for chemoprevention by cyclooxygenase inhibitors.* Cancer 2001, 91: 333-8.
- 44. Takahashi, M., Furukawa, F., Toyoda, K., Sato, H., Hasegawa, R., Imaida, K. et al. *Effects of various prostaglandin synthesis inhibitors on pancreatic carcinogenesis in hamsters after initiation with N-nitrosobis(2-oxopropyl)amine*. Carcinogenesis 1990, 11: 393-5.
- 45. Jaiswal, M., LaRusso, N.F., Gores, G.J. *Nitric oxide in gastrointestinal epithelial cell carcinogenesis: Linking inflammation to oncogenesis.* Am J Physiol Gastrointest Liver Physiol 2001, 281: G626-34.
- 46. Torok, N.J., Higuchi, H., Bronk, S., Gores, G.J. *Nitric oxide inhibits apoptosis downstream of cytochrome c release by nitrosylating caspase 9.* Cancer Res 2002, 62: 1648-53.
- 47. Timoshenko, A.V., Lala, P.K., Chakraborty, C. *PGE2-mediated upregulation of iNOS in murine breast cancer cells through the activation of EP*<sub>4</sub> receptors. Intl J Cancer 2004, 108: 384-9.
- 48. Chan, T.A., Morin, P.J., Vogelstein, B., Kinzler, K.W. *Mechanisms underlying nonsteroidal antiinflammatory drug-mediated apoptosis*. Proc Natl Acad Sci USA 1998, 95: 681-6.
- 49. Key, T.J., Pike, M.C. The role of oestrogens and progestagens in the epidemiology and prevention of breast cancer. Eur J Cancer Clin Oncol 1998, 24: 29-43.
- 50. Thomas, H.V., Key, T.J., Allen, D.S., Moore, J.W., Dowsett, M., Fentiman, I.S., Wang, D.Y. A prospective study of endogenous serum hormone concentrations and breast cancer risk in post-menopausal women on the island of Guernsey. Brit J Cancer 1997, 76: 401-5.
- 51. Helzlsouer, K.J., Alberg, A.J., Bush, T.L., Longcope, C., Gordon, G.B., Comstock, G.W. *A prospective study of endogenous hormones and breast cancer*. Cancer Detect Prev 1994, 18: 79-85.
- 52. Reed, M.J., Owen, A.M., Lai, L.C., Coldham, N.G., Ghilchik, M.W., Shaikh, N.A., James, V.H. *In situ oestrone synthesis in normal breast and breast tumour tissues: Effect of treatment with 4-hydroxyandrostenedione*. Intl J Cancer 1989, 44: 233-7.
- 53. O'Neill, J.S., Elton, R.A., Miller, W.R. *Aromatase activity in adipose tissue from breast quadrants: A link with tumour site.* Brit Med J (Clin Res Ed) 1988, 296: 741-3.
- 54. Zhou, C., Zhou, D., Esteban, J., Murai, J., Siiteri, P.K., Wilczynski, S., Chen, S. *Aromatase gene expression and its exon I usage in human breast tumors. Detection of aromatase messenger RNA by reverse transcription-polymerase chain reaction.* J Steroid Biochem Mol Biol 1996, 59: 163-71.
- 55. Mahendroo, M.S., Mendelson, C.R., Simpson, E.R. *Tissue-specific and hormonally controlled alternative promoters regulate aromatase cytochrome P450 gene expression in human adipose tissue*. J Biol Chem 1993, 268: 19463-70.
- 56. Michael, M.D., Michael, L.F., Simpson, E.R. A CRE-like sequence that binds CREB and contributes to cAMP-dependent regulation of the proximal promoter of the human aromatase P450 (CYP19) gene. Mol Cell Endocrinol 1997, 134: 147-56.

- 57. Zhao, Y., Agarwal, V.R., Mendelson, C.R., Simpson, E.R. Estrogen biosynthesis proximal to a breast tumor is stimulated by PGE2 via cyclic AMP, leading to activation of promoter II of the CYP19 (aromatase) gene. Endocrinology 1996, 137: 5739-42.
- 58. Brueggemeier, R.W., Quinn, A.L., Parrett, M.L., Joarder, F.S., Harris, R.E., Robertson, F.M. *Correlation of aromatase and cyclooxygenase gene expression in human breast cancer specimens.* Cancer Lett 1999, 140: 27-35.
- 59. Jones, M.K., Szabo, I.L., Kawanaka, H., Husain, S.S., Tarnawski, A.S. *Von Hippel Lindau tumor suppressor and HIF-1-new targets of NSAIDs inhibition of hypoxia-induced angiogenesis.* FASEB J 2002, 16: 265-6
- 60. Jaakkola, P., Mole, D.R, Tian, Y.M., Wilson, M.I., Gielbert, J., Gaskell, S.J., von Kriegsheim, A., Hebestreit, H.F., Mukherji, M., Schofield, C.J., Maxwell, P.H., Pugh, C.W, Ratcliffe, P.J. Targeting of HIF- to the von Hippel-Lindau ubiquitination complex by O<sub>2</sub>-regulated prolyl hydroxylation. Science 2001, 292: 468-72
- 61. Ivan, M., Kondo, K., Yang, H., Kim, W., Valiando, J., Ohh, M., Salic, A., Asara, J.M., Lane, W.S., Kaelin, W.G. Jr. *HIF targeted for VHL-mediated destruction by proline hydroxylation: Implications for O<sub>2</sub> sensing.* Science 2001, 292: 464-8.
- 62. Yu, F., White, S.B., Zhao, Q., Lee, F.S. *HIF-1 binding to VHL is regulated by stimulus-sensitive proline hydroxylation*. Proc Natl Acad Sci USA 2001, 98: 9630-5.
- 63. Levy, A.P., Levy, N.S., Goldberg, M.A. *Post-transcriptional regulation of vascular endothelial cell growth factor by hypoxia*. J Biol Chem 1996, 271: 2746-53.
- 64. Gerber, H.J.P., Condorelli, F., Park, J., Ferrara, N. Differential transcriptional regulation of the two vascular endothelial growth factor receptor genes. Flt-1, but not Flk-1/KDR, is up-regulated by hypoxia. J Biol Chem 1997, 272: 23659-67.
- 65. Forsythe, J.A, Jiang, B.-H., Rue, E.A., Semenza, G.L. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular  $O_2$  tension. Proc Natl Acad Sci USA 1995, 92: 5510-4.
- 66. Pugh, C.W., Ratcliffe, P.J. Regulation of angiogenesis by hypoxia: Role of the HIF system. Nat Med 2003, 9: 677-4
- 67. Tarnawski, A.S., Jones, M.K. Inhibition of angiogenesis by NSAIDs: Molecular mechanisms and clinical implications. J Mol Med 2003, 81: 627-36.
- 68. Nishi, H., Nishi, K.H., Johnson, AC. Early growth response-1 gene mediates up-regulation of epidermal growth factor receptor expression during hypoxia. Cancer Res 2002, 62: 827-34.
- 69. Yan, S.F., Fujita, T., Lu, J., Okada, K, Shan Zou, Y., Mackman, N., Pinsky, D.J., Stern, D.M. *Egr-1, a master switch coordinating upregulation of divergent gene families underlying ischemic stress.* Nat Med 2000, 6: 1355-61.
- 70. Stula, M., Orzechowski, H.D., Gschwend, S., Vetter, R., von Harsdorf, R., Dietz, R., Paul, M. *Influence of sustained mechanical stress of Egr-1 mRNA expression in cultured human endothelial cells*. Mol Cell Biochem 2000, 210: 101-8
- 71. Vidal, F., Aragones, J., Alfranca, A., de Landazuri, M.O. *Upregulation of vascular endothelial growth factor receptor Flt-1 after endothelial denudation: Role of transcription factor Egr-1.* Blood 2000, 95: 3387-5.

- 72. Fahmy, R.G., Dass, C.R., Sun, L.Q., Chesterman, C.N., Khachigian, L.M. *Transcription factor Egr-1 supports FGF-dependent angiogenesis during neovascularization and tumor growth.* Nat Med 2003, 9: 1026-32.
- 73. Szabo, I.L, Pai, R., Soreghan, B., Jones, M.K., Baatar, D., Kawanaka, H., Tarnawski, A.S. *NSAIDs inhibit the activation of egr-1 gene in microvascular endothelial cells. A key to inhibition of angiogenesis?* J Physiol (Paris) 2001, 95: 379-83.
- 74. Boon, E.M., Keller, J.J., Wormhoudt, T.A., Giardiello, F.M., Offerhaus, G.J., van der Neut, R., Pals, S.T. Sulindac targets nuclear  $\beta$ -catenin accumulation and Wnt signalling in adenomas of patients with familial adenomatous polyposis and in human colorectal cancer cell lines. Brit J Cancer 2004, 90: 224-9.
- 75. Brown, W.A., Skinner, S.A., Vogiagis, D., O'Brien, P.E. Inhibition of  $\beta$ -catenin translocation in rodent colorectal tumors A novel explanation for the protective effect of nonsteroidal anti-inflammatory drugs in colorectal cancer. Digest Dis Sci 2001, 46: 2314-21.
- 76. Smith, M.L., Hawcroft, G., Hull, M.A. The effect of nonsteroidal anti-inflammatory drugs on human colorectal cancer cells: Evidence of different mechanisms of action. Eur J Cancer 2000, 36: 664-74.
- 77. Dihlmann, S, Klein, S., Doeberitz, M.V. Reduction of  $\beta$ -catenin/T-cell transcription factor signaling by aspirin and indomethacin is caused by an increased stabilization of phosphorylated beta-catenin. Mol Cancer Ther 2003, 2: 509-16.
- 78. Dihlmann, S., Siermann, A., Doeberitz, M.V. *The nonsteroidal anti-inflammatory drugs aspirin and indomethacin attenuate*  $\beta$ -catenin/TCF-4 signaling. Oncogene 2001, 20: 645-53.
- 79. Rice, P.L., Kelloff, J., Sullivan, H., Driggers, L.J., Beard, K.S., Kuwada, S., Piazza, G., Ahnen, D.J. *Sulindac metabolites induce caspase- and proteasome-dependent degradation of*  $\beta$ -catenin protein in human colon cancer cells. Mol Cancer Ther 2003, 2: 885-92.
- 80. Williams, J.L., Borgo, S., Hasan, I., Castillo, E., Traganos, F., Rigas, B. *Nitric oxide-releasing nonsteroidal anti-inflammatory drugs (NSAIDs) alter the kinetics of human colon cancer cell lines more effectively than traditional NSAIDs: Implications for colon cancer chemoprevention.* Cancer Res 2001, 61: 3285-9.
- 81. Williams, J.L., Nath, N., Chen, J., Hundley, T.R., Gao, J., Kopelovich, L., Kashfi, K., Rigas, B. *Growth inhibition of human colon cancer cells by nitric oxide (NO)-donating aspirin is associated with cyclooxygenase-2 induction and \beta-catenin/T-cell factor signaling, nuclear factor-\kappaB, and NO synthase 2 inhibition: Implications for chemoprevention. Cancer Res 2003, 63: 7613-8.*
- 82. Nath, N., Kashfi, K., Chen, J., Rigas, B. *Nitric oxide-donating aspirin inhibits beta-catenin T cell factor (TCF) signaling in SW480 colon cancer cells by disrupting the nuclear*  $\beta$ -catenin-TCF association. Proc Natl Acad Sci USA 2003, 100: 12584-9.
- 83. Zhou, X.M., Wong, B.C.Y., Fan, X.M., Zhang, H.B., Lin, M.C.M., Kung, H.F., Fan, D.M., Lam, S.K. *Non-steroidal anti-inflammatory drugs induce apoptosis in gastric cancer cells through up-regulation of bax and bak*. Carcinogenesis 2001, 22: 1393-7.
- 84. Lim, J.T.E., Piazza, G.A., Han, E.K.H, Delohery, T.M., Li, H., Finn, T.S., Buttyan, R., Yamamoto, H., Sperl, G.J., Brendel, K., Gross, P.H., Pamukcu, R., Weinstein, I.B. *Sulindac derivatives*

- inhibit growth and induce apoptosis in human prostate cancer cell lines. Biochem Pharmacol 1999, 58: 1097-107.
- 85. Liu, X.H., Yao, S., Kirschenbaum, A., Levine, A.C. NS398, a selective cyclooxygenase-2 inhibitor, induces apoptosis and down-regulates bcl-2 expression in LNCaP cells. Cancer Res 1998, 58: 4245-9.
- 86. Dormond, O., Ruegg, C. Inhibition of tumor angiogenesis by non-steroidal anti-inflammatory drugs: Emerging mechanisms and therapeutic perspectives. Drug Resist Updates 2001, 4: 314-21.
- 87. Hsu, A.L., Ching, T.T., Wang, D.S., Song, X.Q., Rangnekar, V.M., Chen, C.S. *The cyclooxygenase-2 inhibitor celecoxib induces apoptosis by blocking Akt activation in human prostate cancer cells independently of Bcl-2.* J Biol Chem 2000, 275: 11397-403.
- 88. Berman, K.S., Verma, U.N., Harburg, G., Minna, J.D., Cobb, M.H., Gaynor, R.B. *Sulindac enhances tumor necrosis factor-\alpha-mediated apoptosis of lung cancer cell lines by inhibition of nuclear factor-\kappaB. Clin Cancer Res 2002, 8: 354-60.*
- 89. Jones, M.K., Wang, H.T., Peskar, B.M., Levin, E., Itani, R.M., Sarfeh, I.J., Tarnawski, A.S. *Inhibition of angiogenesis by non-steroidal anti-inflammatory drugs: Insight into mechanisms and implications for cancer growth and ulcer healing.* Nat Med 1999, 5: 1418-23.
- 90. Shureiqi, I., Jiang, W., Fischer, S.M., Xu, X.C., Chen, D.N., Lee, J.J., Lotan, R., Lippman, S.M. *GATA-6 transcriptional regulation of 15-lipoxygenase-1 during NSAID-induced apoptosis in colorectal cancer cells.* Cancer Res 2002, 62: 1178-83.
- 91. Shappell, S.B., Manning, S., Boeglin, W.E., Guan, Y.F., Roberts, R.L., Davis, L., Olson, S.J., Jack, G.S., Coffey, C.S., Wheeler, T.M., Breyer, M.D., Brash, A.R. *Alterations in lipoxygenase and cyclooxygenase-2 catalytic activity and mRNA expression in prostate carcinoma*. Neoplasia 2001, 3: 287-303.
- 92. Shureiqi, I., Chen, D., Lotan, R., Yang, P., Newman, R.A., Fischer, S.M., Lippman, S.M. 15-Lipoxygenase-1 mediates non-steroidal anti-inflammatory drug-induced apoptosis independently of cyclooxygenase-2 in colon cancer cells. Cancer Res 2000, 60: 6846-50.
- 93. Shureiqi, I., Chen, D., Lee, J.J., Yang, P., Newman, R.A., Brenner, D.E., Lotan, R., Fischer, S.M., Lippman, S.M. *15-LOX-1: A novel molecular target of nonsteroidal anti-inflammatory drug-induced apoptosis in colorectal cancer cells.* J Natl Cancer Inst 2000, 92: 1136-42.
- 94. Shureiqi, I., Xu, X., Chen, D., Lotan, R., Morris, J.S., Fischer, S.M., Lippman, S.M. *Nonsteroidal anti-inflammatory drugs induce apoptosis in esophageal cancer cells by restoring 15-lipoxygenase-1 expression.* Cancer Res 2001, 61: 4879-84.
- 95. Xu, X.C., Shappell, S.B., Liang, Z., Song, S., Menter, D., Subbarayan, V., Iyengar, S., Tang, D.G., Lippman, S.M. Reduced 15S-lipoxygenase-2 expression in esophageal cancer specimens and cells and upregulation in vitro by the cyclooxygenase-2 inhibitor, NS398. Neoplasia 2003, 5: 121-7.
- 96. Wu, J., Xia, H.H., Tu, S.P., Fan, D.M., Lin, M.C., Kung, H.F., Lam, S.K., Wong, B.C. *15-Lipoxygenase-1 mediates cyclooxygenase-2 inhibitor-induced apoptosis in gastric cancer.* Carcinogenesis. 2003, 24: 243-7

- 97. Shureiqi, I., Jiang, W., Zuo, X., Wu, Y., Stimmel, J.B., Leesnitzer, L.M., Morris, J.S., Fan, H.Z., Fischer, S.M, Lippman, S.M. *The 15-lipoxygenase-1 product 13-S-hydroxyoctadecadienoic acid down-regulates PPAR-δ to induce apoptosis in colorectal cancer cells.* Proc Natl Acad Sci USA 2003, 100: 9968-73.
- 98. Hao, C.M., Redha, R., Morrow, J., Breyer, M.D. *Peroxisome* proliferator-activated receptor  $\delta$  activation promotes cell survival following hypertonic stress. J Biol Chem 2002, 277: 21341-5.
- 99. Glinghammar, B., Skogsberg, J., Hamsten, A., Ehrenborg, E. *PPAR*  $\delta$  activation induces *COX-2* gene expression and cell proliferation in human hepatocellular carcinoma cells. Biochem Biophys Res Comm 2003, 308: 361-8.
- 100. Pan, Y.Q., Zhang, J.S., Gazi, M.H., Young, C.Y.F. *The cyclooxygenase 2-specific nonsteroidal anti-inflammatory drugs celecoxib and nimesulide inhibit androgen receptor activity via induction of c-Jun in prostate cancer cells*. Cancer Epidemiol Biomark Prevent 2003, 12: 769-74.
- 101. Zhu, W., Smith, A., Young, C.Y.F. A nonsteroidal anti-inflammatory drug, flufenamic acid, inhibits the expression of the androgen receptor in LNCaP cells. Endocrinology 1999, 140: 5451-4.
- 102. Brawer, M.K., Ellis, W.J. Chemoprevention for prostate cancer. Cancer 1995, 75: 1783-9.
- 103. Pienta, K., Esper, P. *Risk factors for prostate cancer*. Ann Intern Med 1993, 118: 793-803.
- 104. Kolonel, L.N., Nomura, A.M.Y., Cooney, R.V. *Dietary fat and prostate cancer: Current status.* J Natl Cancer Inst 1999, 91: 414-28.
- 105. Dorgan, J.F., Judd, J.T., Longcope, C., Brown, C., Schatzkin, A., Clevidence, B.A., Campbell, W.S., Nair, P.P., Franz, C., Kahle, L., Taylor, P.R. Effects of dietary fat and fiber on plasma and urine androgens and estrogens in men A controlled feeding study. Am J Clin Nutr 1996, 64: 850-5.
- 106. Hamalainen, E., Adlercreutz, H., Puska, P., Pietinen, P. *Diet and serum sex hormones in healthy men.* J Steroid Biochem 1984, 20: 459-64.
- 107. Gann, P.H., Hennekens, C.H., Ma, J., Longcope, C., Stampfer, M.J. *Prospective study of sex hormone levels and risk of prostate cancer.* J Natl Cancer Inst 1996, 88: 1118-26.
- 108. Lane, K.E., Leav, I., Ziar, J., Bridges, R.S., Rand, W.M., Ho, S.M. Suppression of testosterone and estradiol- $17\beta$ -induced dysplasia in the dorsolateral prostate of Noble rats by bromocriptine. Carcinogenesis 1997, 18: 1505-10.
- 109. Yaono, M., Tamano, S., Mori, T., Kato, K., Imaida, K., Asamoto, M., Shirai, T. *Lobe specific effects of testosterone and estrogen on 3,2'-dimethyl-4-aminobiphenyl-induced rat prostate carcinogenesis*. Cancer Lett 2000, 150: 33-40.
- 110. Lindzey, J., Kumar, M.V., Grossmann, M., Young, C.Y.F., Tindall, D.J. *Molecular mechanisms of androgen action.* Vitamin Horm 1994, 49: 383-432.
- 111. Thompson, L., Feigl, P., Coltman, C. Chemoprevetion of prostate cancer with finasteride. In: Important Advances in Oncology. V.T. DeVita, Jr., S. Hellman, S.A.Rosenberg (Eds.). J.B. Lippincott: Philadelphia, 1996, 57-76.

- 112. Simard, J., Dumont, M., Soucy, P., Labrie, F. *Perspective: Prostate cancer susceptibility genes.* Endocrinology 2002, 143: 2029-40.
- 113. Hayes, R.B. *Gene-environment interrelations in prostate cancer*. Epidemiol Rev 2001, 23: 163-7.
- 114. Lieberman, R., Bermejo, C., Akaza, H., Greenwald, P., Fair, W., Thompson, I. *Progress in prostate cancer chemoprevention: Modulators of promotion and progression.* Urology 2001, 58: 835-42.
- 115. Ripple, M.O., Henry, W.F., Schwarze, S.R., Wilding, G., Weindruch, R. *Effect of antioxidants on androgen-induced AP-1 and NF-κB DNA-binding activity in prostate carcinoma cells.* J Natl Cancer Inst 1999, 91: 1227-32.
- 116. Fleshner, N.E., Klotz, L.H. *Diet, androgens, oxidative stress and prostate cancer susceptibility.* Cancer Metast Rev 1998, 17: 325-30.
- 117. Truica, C.I., Byers, S., Gelmann, E.P.  $\beta$ -Catenin affects androgen receptor transcriptional activity and ligand specificity. Cancer Res 2000, 60: 4709-13.
- 118. Lu, M.L., Schneider, M.C., Zheng, Y.X., Zhang, X.B., Richie, J.P. *Caveolin-1 interacts with androgen receptor A positive modulator of androgen receptor mediated transactivation.* J Biol Chem 2001, 276: 13442-51.
- 119. Gregory, C.W., He, B., Johnson, R.T., Ford, O.H., Mohler, J.L., French, F.S., Wilson, E.M. *A mechanism for androgen receptor-mediated prostate cancer recurrence after androgen deprivation therapy.* Cancer Res 2001, 61: 4315-9.
- 120. Chesire, D.R., Isaacs, W.B. Ligand-dependent inhibition of  $\beta$ -catenin/TCF signaling by androgen receptor. Oncogene 2002, 21: 8453-69.
- 121. Song, L.N., Herrell, R., Byers, S., Shah, S., Wilson, E.M., Gelmann, E.P.  $\beta$ -Catenin binds to the activation function 2 region of the androgen receptor and modulates the effects of the N-terminal domain and TIF2 on ligand-dependent transcription. Mol Cell Biol 2003, 23: 1674-87.
- 122. Yang, F.J., Li, X.Y., Sharma, M., Sasaki, C.Y., Longo, D.L., Lim, B., Sun, Z.J. *Linking*  $\beta$ -catenin to androgen-signaling pathway. J Biol Chem 2002, 277: 11336-44.
- 123. Yamamoto, A., Hashimoto, Y., Kohri, K., Ogata, E., Kato, S., Ikeda, K., Nakanishi, M. *Cyclin E as a coactivator of the androgen receptor.* J Cell Biol 2000, 150: 873-9.
- 124. Hayes, S.A., Zarnegar, M., Sharma, M., Yang, F.J., Peehl, D.M., ten Dijke, P., Sun, Z.J. *SMAD3 represses androgen receptor-mediated transcription*. Cancer Res 2001, 61(5): 2112-8.
- 125. Kang, H.Y., Huang, K.E., Chang, S.Y., Ma, W.L., Lin, W.J., Chang, C. *Differential modulation of androgen receptor-mediated transactivation by Smad3 and tumor suppressor Smad4*. J Biol Chem 2002, 277: 43749-56.
- 126. Poukka, H., Aarnisalo, P., Santti, H., Janne, O.A., Palvimo, J.J. Coregulator small nuclear RING finger protein (SNURF) enhances Sp1- and steroid receptor-mediated transcription by different mechanisms. J Biol Chem 2000, 275: 571-9.
- 127. Petre, C.E., Wetherill, Y.B., Danielsen, M., Knudsen, K.E. *Cyclin D1: Mechanism and consequence of androgen receptor co-repressor activity.* J Biol Chem 2002, 277: 2207-15.

- 128. Kotaja, N., Aittomaki, S., Silvennoinen, O., Palvimo, J.J., Janne, O.A. *ARIP3* (androgen receptor-interacting protein 3) and other PIAS (protein inhibitor of activated STAT) proteins differ in their ability to modulate steroid receptor-dependent transcriptional activation. Mol Endocrinol 2000, 14: 1986-2000.
- 129. Liao, G.Q., Chen, L.Y, Zhang, A.H., Godavarthy, A., Xia, F., Ghosh, J.C., Li, H., Chen, J.D. *Regulation of androgen receptor activity by the nuclear receptor corepressor SMRT*. J Biol Chem 2003, 278: 5052-61.
- 130. Irvine, R.A., Yu, M.C., Ross, R.K., Coetzee, G.A. *The CAG and GCC micro-satellites of the androgen receptor gene are in linkage disequilibrium in men with prostate cancer.* Cancer Res 1995, 55: 1937-40.
- 131. Hsing, A.W., Gao, Y.T., Wu, G., Wang, X., Deng, J., Chen, Y.L., Sesterhenn, I.A., Mostofi, F.K., Benichou, J., Chang, C.S. Polymorphic CAG and GGN repeat lengths in the androgen receptor gene and prostate cancer risk: A population-based case-control study in China. Cancer Res 2000, 60: 5111-6.
- 132. Platz, E.A., Rimm, E.B., Willett, W.C., Kantoff, P.W., Giovanucci, E. *Racial variation in prostate cancer incidence and in the hormonal system markers among male health professionals.* J Natl Cancer Inst 2000, 92: 2009-17.
- 133. Zha, S., Gage, W.R., Sauvageot, J., Saria, E.A., Putzi, M.J., Ewing, C.M., Faith, D.A., Nelson, W.G., De Marzo, A.M., Isaacs, W.B. *Cyclooxygenase-2 is up-regulated in proliferative inflammatory atrophy of the prostate, but not in prostate carcinoma*. Cancer Res 2001, 61: 8617-23
- 134. Wagner, E.F. *AP-1 Introductory remarks*. Oncogene 2001, 20: 2334-5.
- 135. Hayes, J.D., McMahon, M. *Molecular basis for the contribu*tion of the antioxidant responsive element to cancer chemoprevention. Cancer Lett 2001, 174: 103-13.
- 136. Kataoka, K., Shioda, S., Yoshitomo-Nakagawa, K., Handa, H., Nishizawa, M. *Maf and Jun nuclear oncoproteins share downstream target genes for inducing cell transformation.* J Biol Chem 2001, 276: 36849-56.
- 137. Lee, S.K., Kim, J.H., Lee, Y.C., Cheong, J.H., Lee, J.W. Silencing mediator of retinoic acid and thyroid hormone receptors, as a novel transcriptional corepressor molecule of activating protein-1, nuclear factor-κB, and serum response factor. J Biol Chem 2000, 275: 12470-4.
- 138. Rogatsky, I., Zarember, K.A., Yamamoto, K.R. Factor recruitment and TIF2/GRIP1 corepressor activity at a collagenase-3 response element that mediates regulation by phorbol esters and hormones. EMBO J 2001, 20: 6071-83.
- 139. Suzukawa, K., Colburn, N.H. *AP-1 transrepressing retinoic acid does not deplete coactivators or AP-1 monomers but may target specific Jun or Fos containing dimers.* Oncogene 2002, 21: 2181-90.
- 140. Pessah, M., Prunier, C., Marais, J., Ferrand, N., Mazars, A., Lallemand, F., Gauthier, J.M., Afti, A. *c-Jun interacts with the corepressor TG-interacting factor (TGIF) to suppress Smad2 transcriptional activity.* Proc Natl Acad Sci USA 2001, 98: 6198-203.
- 141. Sharma, M., Sun, Z. 5 'TG3' interacting factor interacts with Sin3A and represses AR-mediated transcription. Mol Endocrinol 2001, 15: 1918-28.

- 142. Shabsigh, A., Ghafar, M.A., de la Taille, A., Burchardt, M., Kaplan, S.A., Anastasiadis, A.G., Buttyan, R. *Biomarker analysis demonstrates a hypoxic environment in the castrated rat ventral prostate gland*. J Cell Biochem 2001, 81: 437-44.
- 143. Feng, Z., Joos, H.J., Vallan, C., Muhlbauer, R., Altermatt, H.J., Jaggi, R. *Apoptosis during castration-induced regression of the prostate is Fos dependent.* Oncogene 1998, 17: 2593-600.
- 144. Murtha, P.E., Zhu, W., Zhang, J., Zhang, S., Young, C.Y. Effects of Ca<sup>2+</sup> mobilization on expression of androgen-regulated genes: Interference with androgen receptor-mediated transactivation by AP-I proteins. Prostate 1997, 33: 264-70.
- 145. Chung, B.H., Mitchell, S.H., Zhang, J.S., Young, C.Y. Effects of docosahexaenoic acid and eicosapentaenoic acid on androgen-mediated cell growth and gene expression in LNCaP prostate cancer cells. Carcinogenesis 2001, 22: 1201-6.
- 146. Yang-Yen, H.F., Chambard, J.C., Sun, Y.L., Smeal, T., Schmidt, T.J., Drouin, J., Karin, M. *Transcriptional interference between c-Jun and the glucocorticoid receptor: Mutual inhibition of DNA binding due to direct protein-protein interaction.* Cell 1990, 62: 1205-15.
- 147. Schule, R., Rangarajan, P., Kliewer, S., Ransone, L.J., Bolado, J., Yang, N., Verma, I.M., Evans, R. M. Functional antagonism between oncoprotein c-Jun and the glucocorticoid receptor. Cell 1990, 62: 1217-26.
- 148. Jonat, C., Rahmsdorf, H.J., Park, K.K., Cato, A.C., Gebel, S., Ponta, H., Herrlich, P. *Antitumor promotion and antiinflammation: Down-modulation of AP-1 (Fos/Jun) activity by glucocorticoid hormone.* Cell 1990, 62: 1189-204.
- 149. Tzukerman, M., Zhang, X.K., Pfahl, M. Inhibition of estrogen receptor activity by the tumor promoter 12-O-tetradeconylphorbol-13-acetate: A molecular analysis. Mol Endocrinol 1991, 5: 1983-92.
- 150. Jorgensen, J.S., Nilson, J.H. AR suppresses transcription of the  $\alpha$  glycoprotein hormone subunit gene through protein-protein interactions with cJun and activation transcription factor 2. Mol Endocrinol 2001, 15: 1496-504.

- 151. Wang, Q., Lu, J.H., Yong, E.L. Ligand- and coactivator-mediated transactivation function (AF2) of the androgen receptor ligand-binding domain is inhibited by the cognate hinge region. J Biol Chem 2001, 276: 7493-9.
- 152. Aarnisalo, P., Palvimo, J.J., Janne, O.A. *Creb-binding protein in androgen receptor-mediated signaling*. Proc Natl Acad Sci USA 1998, 95: 2122-7.
- 153. Sato, N., Sadar, M.D., Bruchovsky, N., Saatcioglu, F., Rennie, P.S., Sato, S., Lange, P.H., Gleave, M E. *Androgenic induction of prostate-specific antigen gene is repressed by protein-protein interaction between the androgen receptor and AP-1/c-Jun in the human prostate cancer cell line LNCaP.* J Biol Chem 1997, 272: 17485-94.
- 154. Fronsdal, K., Engedal, N., Slagsvold, T., Saatcioglu, F. *CREB binding protein is a coactivator for the androgen receptor and mediates cross-talk with AP-1*. J Biol Chem 1998, 273: 31853-9.
- 155. Rigas, B., Shiff, S.J. Is inhibition of cyclooxygenase required for the chemopreventive effect of NSAIDs in colon cancer? A model reconciling the current contradiction. Med Hypotheses 2000, 54: 210-5.
- 156. Wechter, W.J., Leipold, D.D., Murray, E.D., Quiggle, D., McCracken, J.D., Barrios, R.S., Greenberg, N.M. *E-7869 (R-flur-biprofen) inhibits progression of prostate cancer in the TRAMP mouse.* Cancer Res 2000, 60: 2203-8.
- 157. Wechter, W.J., Murray, E.D., Kantoci, D., Quiggle, D.D., Leipold, D.D., Gibson, K.M., McCracken, J.D. *Treatment and survival study in the C57BL/6J-APC(Min)/+ (Min) mouse with R-flurbiprofen.* Life Sci 2000, 66: 745-53.
- 158. Raz, A. Is inhibition of cyclooxygenase required for the anti-tumorigenic effects of nonsteroidal, anti-inflammatory drugs (NSAIDs)? In vitro versus in vivo results and the relevance for the prevention and treatment of cancer. Biochem Pharmacol 2002, 63: 343-7.